

## Protein extraction from Tissues and Cultured Cells using Bioruptor® Standard & Plus

### Introduction

Protein extraction from tissues and cultured cells is the first step for many biochemical and analytical techniques (PAGE, Western blotting, mass spectrometry, etc.) or protein purification. Efficient disruption and homogenization of animal tissues and cultured cells are required to ensure high yields of proteins. Diagenode's Bioruptor® uses state-of-the-art ultrasound technology to efficiently disrupt and homogenize tissues and cultured cells in just one step. Bioruptor® offers unique benefits for tissue disruption and homogenization:

- Fast and simple
- No contamination between samples
- Efficient
- Gentle processing
- Reproducible
- Temperature controlled
- Multiplexing capability

### General remarks before starting

- Conditions for protein extraction (e.g. use of fresh or frozen tissue, composition of extraction buffer etc.) must be adjusted according to the nature of the proteins of interest and the assays to be run. SDS might be added to the extraction buffer to maximize the yield of soluble proteins. SDS extracts can be used for SDS electrophoresis and Western blotting. It is recommended to reduce the SDS concentration for 2D electrophoresis, enzyme-linked immunosorbent assay and mass spectrometry.
- For functional studies (e.g. the study of protein-protein interactions), avoid using ionic detergents and high concentrations of salt.

**Extraction buffer:** use RIPA buffer as a starting point for optimization:

50 mM Tris-HCl (pH 7.4)

150 mM NaCl

1% NP-40

0.25% Na-deoxycholate

Protease Inhibitor Mix

SDS 0.1 - 2% (optional)

*It is always recommended to optimize the buffer composition depending on a specific research project*

- Always use Protease Inhibitor Mix during extraction procedure to block the possible protein degradation.

- Use Diagenode's TPX tubes for sonication. Depending on the desired final volume, 1.5 ml TPX microtubes (Cat. No.: C30010010-50 or C30010010-1000) or 15 ml TPX tubes (Diagenode, Cat. No.: C30010009) might be used. Always respect the recommended sonication volumes: 100 - 300 µl for 1.5 ml TPX tubes and 1 - 2 ml for 15 ml TPX tubes (strictly follow the Bioruptor® instructions as shown in the corresponding manual before starting any sonication experiments).
- Keep extracted proteins at -80°C.

## Required materials and reagents

- Bioruptor® Standard (Cat. No. B01010001) or Plus (Cat. No. B01020001)
- Water Cooler (Cat. No. B02010003; 115V or B02010002; 230 V)
- Single Cycle Valve (Cat. No. B02020004) (required for Bioruptor® Plus)
- Tube holder for 1.5 ml tubes (Cat. No. B01200011)
- 15 ml sonication accessories for Bioruptor® Standard & Plus (Cat. No. B01200015)
- Choose between option **A** or **B**:  
**A<sub>1</sub>**: 1.5 ml (Cat. No. C30010010-50 or C30010010-1000) or 15 ml TPX tubes (Cat. No. C30010009) for sonication  
**A<sub>2</sub>**: Protein Extraction Beads (Cat. No. C20000021) for tissue disruption (**not required for cell lysis**)  
**B**: Protein Extraction kit (Cat. No. C20000020); pre-filled 15 ml TPX tubes
- Protease Inhibitor Mix (Cat. No. C12010011 or C12010012)
- Buffer for protein extraction from tissue or cell lysis (not supplied)
- Reagents for protein quantification (optional)

## Protocol

### I. Protein extraction from Tissues

- » *This protocol has been validated for up to 50 mg of tissue. Do not use more tissue per sample. For larger quantity cut the tissue and proceed to the disruption in separate tubes. When proceeding 20 - 50 mg of tissue 15 ml TPX tubes are recommended with a final volume of 1 - 2 ml. Less tissue could be sonicated in 1.5 ml TPX tubes with a final volume of 100 - 300 µl.*
- » *Minimize the time of tissue collection to prevent protein degradation.*
- » *Dissected tissues can be snap-frozen in liquid nitrogen and stored at -80°C until protein extraction*

1. Pre-cool Bioruptor® to 4°C using the Bioruptor® Water Cooler (Diagenode, Cat. No. BioAcc-Cool).
2. Fill the TPX tubes with Protein Extraction Beads.
  - » *The recommended quantity of the beads is 200 - 250 mg for 15 ml TPX tubes, 40 - 50 mg for 1.5 ml TPX tubes.*



**Note: If using pre-filled tubes (Cat. No. C20000020 Protein Extraction kit) please skip this step!**

3. Add Protease Inhibitor Mix (200x) to the cold protein extraction buffer: 5 µl per 1 ml of extraction buffer. Scale accordingly.

4. Add the required volume of a cold extraction buffer to the TPX tubes filled with Protein Extraction Beads.
5. Add tissue pieces to the TPX tubes. Make sure that the final volume is in the recommended range: 100 - 300 µl for 1.5 ml TPX microtubes and 1 - 2 ml for 15 ml TPX tubes.
6. Vortex tubes briefly and proceed to sonication by using the Bioruptor® with the following settings:  
*Power: H position (High)*  
*Sonication cycle: 30 sec ON/30 sec OFF*  
*Total sonication time: 5 - 15 cycles*  
*Temperature: 4°C*
  - » *To guarantee homogeneity of sonication, the tube holder should be always completely filled with tubes.*
7. Stop the Bioruptor® after each 5 cycles, vortex samples and check the sample visually for disruption.
  - » *Please note that the optimization might be required depending on the sample format (fresh or frozen tissue), tissue type and tissue amount. The shortest sonication time should be chosen to prevent protein damage. Incomplete disruption may occur with fibrous tissues (i.e. muscles).*
8. Transfer the supernatant to a new tube and centrifuge samples at 14,000 rpm for 15 min at 4°C to remove any remaining insoluble material.
  - » *The Protein Extraction Beads might be washed once with extraction buffer for maximum recovery of total protein but this will lead to the sample dilution.*
9. Transfer the supernatant containing soluble proteins to a new tube.
10. Take an aliquot for the quantification and the further analysis if needed. Store proteins extracts in small aliquots at -80°C.
  - » *Different protein concentration assays exist including: absorbance at 280 nm, Lowry Assay, Bradford Assay, Bicinchoninic Assay (BCA) etc.. Many commercial kits for protein quantification are also available. Please note that measuring the protein concentration in an SDS extract requires that the assay is compatible with the detergent and reducing agent in the solution.*

## **II. Protein extraction from Cultured Cells**

- » *This protocol has been validated using RIPA buffer but it may be necessary to optimize the buffer composition depending on a specific research project.*
  - » *We recommend using 100 µl of an appropriate lysis buffer per  $1 \times 10^6$  cells.*
  - » *For Western blotting, cells might be lysed directly in 1x Laemmli buffer. After sonication, centrifuge extract at 14,000 rpm for 15 min. Transfer the supernatant to a new tube and boil for 3 min. The supernatant can be used in Western blot. Note that protein quantification by common methods is not compatible with Laemmli buffer.*
1. Pre-cool Bioruptor® to 4°C using the Water Cooler.
  2. Add Protease Inhibitor Mix (200x) to the ice-cold cell lysis buffer: 5 µl per 1 ml of extraction buffer. Scale accordingly.

**3. For monolayer cells:**

Rinse the monolayer cells 3 times with cold PBS. For the final rinse, use a cell scraper and transfer the cell suspension to a TPX tube. Centrifuge cells at 1,500 rpm for 10 min at 4°C and aspirate as much supernatant as possible. Proceed to step 4.

**For suspension cells:**

Centrifuge suspension at 1,500 rpm for 10 min at 4°C and aspirate the supernatant. Resuspend the pellet in cold PBS, transfer to a TPX tube and centrifuge at 1,500 rpm for 10 min at 4°C. Aspirate the supernatant. Repeat 2 more times. Proceed to the step 4.

**4. Add ice-cold cell lysis buffer and resuspend the pellet. Incubate on ice for 10 min.**

» *The viscosity may appear at this step*

**5. Vortex tubes briefly and proceed to sonication by using the Bioruptor® with the following settings:**

*Power: H position (High)*

*Sonication cycle: 30 sec ON/30 sec OFF*

*Total sonication time: 5-10 cycles*

*Temperature: 4°C*

» *To guarantee homogeneity of sonication, the tube holder should be always completely filled with tubes.*

**6. Stop the Bioruptor® after 5 cycles, briefly vortex samples and visually check the samples: Samples should be in solution (viscosity should be reduced)**

» *Please note that the optimization might be required depending on sample format (cell density, cell type etc.). The shortest sonication time should be chosen to prevent protein damage.*

**7. Transfer the supernatant to a new tube and centrifuge samples at 14,000 rpm for 15 min at 4°C to remove any remaining insoluble material.**

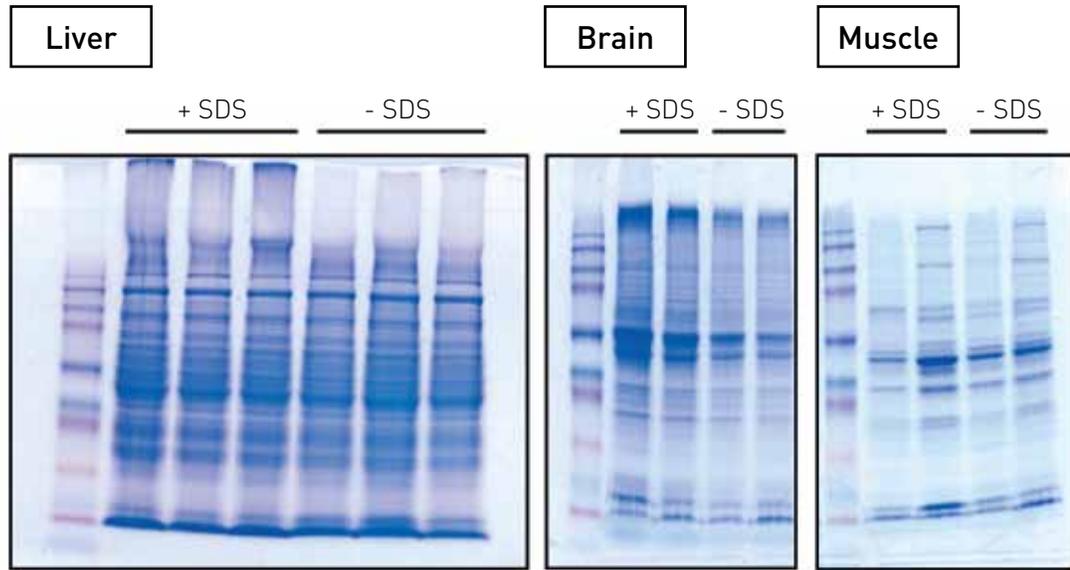
**8. Take an aliquot for the quantification and the further analysis if needed. Store protein extracts at -80°C.**

» *Different protein concentration assays exist including: absorbance at 280 nm, Lowry Assay, Bradford Assay, Bicinchoninic Assay (BCA) etc. Many commercial kits for protein quantification are also available. Please note that measuring the protein concentration in an SDS extract requires that the assay is compatible with the detergent and reducing agent in the solution.*



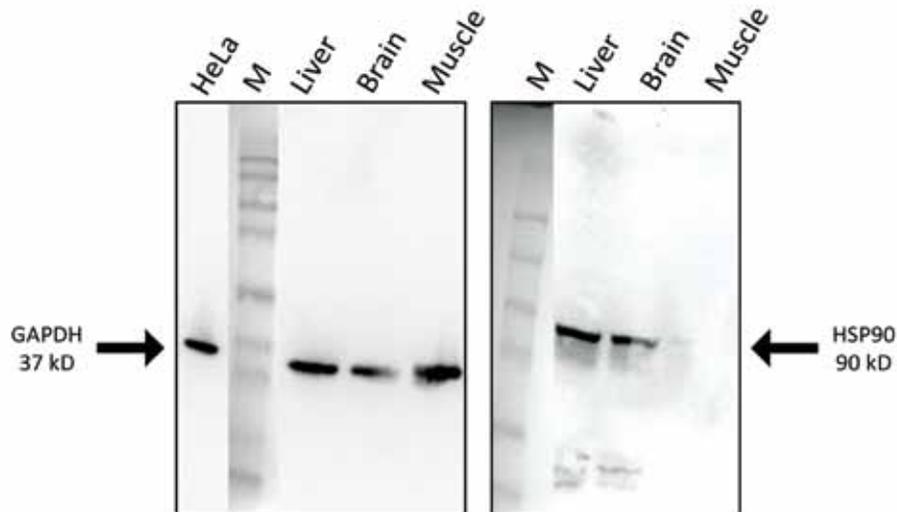
**Figure 1. Protein Extraction Beads are required for efficient tissue disruption using the Bioruptor® Plus**

Complete disruption is observed in the sample containing Diagenode's Protein Extraction Beads (left) after 5 cycles while non-disrupted tissue is still present in the sample without the Protein Extraction Beads (right).



**Figure 2. Total proteins effectively extracted from tissues using Bioruptor® Plus**

Various mouse tissues were disrupted in RIPA buffer supplemented with or without 2% SDS. Total proteins were separated by SDS-PAGE and stained with Coomassie Blue dye.



**Figure 3. Western blot analysis of GAPDH and HSP90 proteins in tissue and cultured cell extracts.**

Expected bands of 37 kD and 90 kD are observed for GAPDH (left panel) and HSP90 (right panel), respectively, in liver, brain and skeletal muscle. Note that HSP90 is expressed in muscle in an extremely low level (H. Quraishi and I. R. Brown, *J Neurosci Res.* 1996 Feb 1;43(3):335-45). Whole cell extract from HeLa cells is loaded as positive control. HeLa cells were lysed using the Bioruptor®.