diagenode

Innovating Epigenetic Solutions

Premium WGBS Kit

Whole Genome Bisulfite Sequencing

Cat. No. C02030034 (8 rxns)

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Contacts

DIAGENODE HEADQUARTERS

Diagenode s.a. BELGIUM | EUROPE
LIEGE SCIENCE PARK
Rue Bois Saint-Jean, 3
4102 Seraing - Belgium
Tel: +32 4 364 20 50
Fax: +32 4 364 20 51
orders@diagenode.com
info@diagenode.com

Diagenode Inc. USA | NORTH AMERICA
400 Morris Avenue, Suite #101
Denville, NJ 07834
Tel: +1 862 209-4680
Fax: +1 862 209-4681
orders.na@diagenode.com
info.na@diagenode.com

For a complete listing of Diagenode's international distributors visit:
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Diagenode website: www.diagenode.com

Content

Introduction

Whole Genome Bisulfite Sequencing (WGBS) is a single nucleotide resolution technique that allows the user to study DNA methylation sites and their role in gene regulation. The Diagenode's **Premium WGBS Kit** is designed to prepare single and paired-end bisulfite converted DNA libraries for sequencing using Illumina® platforms. It has also been validated for the preparation of bisulfite converted library from ChIP'd samples in order to perform **ChIP-Bis-Sequencing**. For Reduced Representation Bisulfite Sequencing (RRBS) experiments use the Diagenode's Premium RRBS Kit (Cat. No.C02030032 and C02030033).

The Diagenode's Premium WGBS Kit contains specially designed enzymes and buffers needed for genome wide bisulfite sequencing. This kit includes:

- a highly efficient adapter ligation step which results in superior library preparations and a maximum number of unique sequencing reads
- a specially designed WGBS enzymatic ligation mix which allows users to perform ligations with longer adapters and better ligation efficiencies
- the MethylTaq Plus 2X Master Mix, a robust polymerase designed to handle bisulfite converted DNA.

These optimizations permit to start a WGBS experiment with amounts of DNA as low as **5 ng**.

Moreover, the Premium WGBS Kit simplifies the workflow by using master mixed reagents and magnetic bead based cleanup, reducing pipetting and eliminating time consuming steps in library preparation.

Kit Method Overview and Time Table

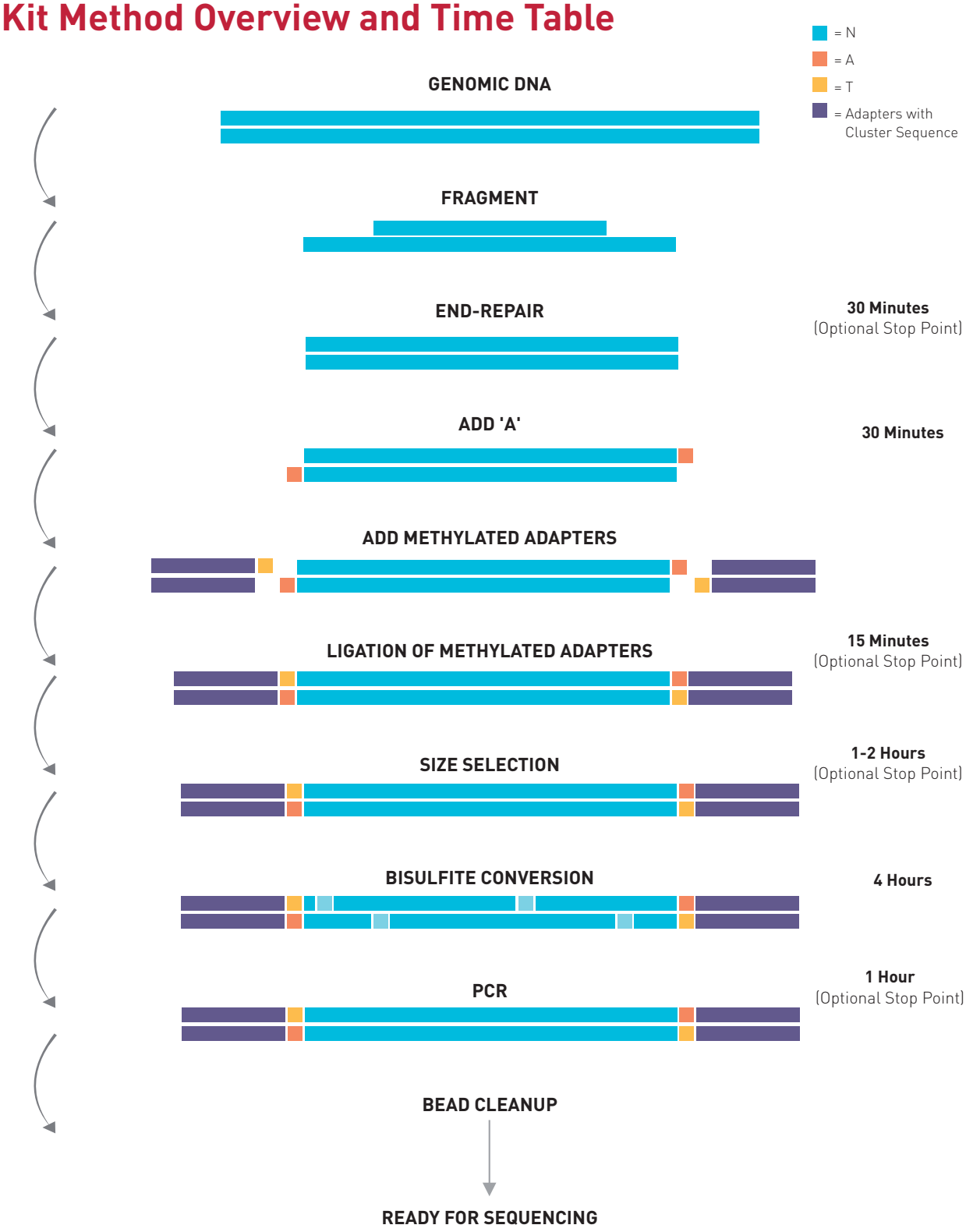

Figure 1: Flow chart for sample preparation

Kit Materials

The Premium WGBS Kit contains enough material to prepare 8 genomic DNA samples for Illumina® compatible sequencing. The shelf life of all reagents is 6 months when stored properly.

Store the components at the indicated temperature upon receipt.

Table 1. Components supplied with the Premium WGBS Kit

Description	Quantity	Storage
Water (white)	1.5 mL	-20°C
WGBS End Repair Buffer Mix (clear)	56 µL	-20°C
WGBS End Repair Enzyme Mix (clear)	24 µL	-20°C
WGBS Adenylation Mix (red)	36 µL	-20°C
WGBS Ligation Mix (purple)	252 µL	-20°C
WGBS Adapter (purple)	20 µL	-20°C
WGBS Adapter Dilution Buffer (purple)	500 µL	-20°C
6x Loading Dye (orange)	500 µL	-20°C
MW Ladder Ready-to-Load 100 bp (orange)	80 µL	-20°C
Resuspension Buffer (white)	2 mL	-20°C
Column Elution Buffer (yellow)	1.8 mL	-20°C
DNA Binding Buffer	24 mL	Room temperature
DNA Wash Buffer	3 mL	Room temperature
Clean-Up Spin Columns	8 columns with tubes	Room temperature
BS Conversion reagent	1 tube	Room temperature
BS Dilution Buffer (black)	300 µL	Room temperature
BS Solubilization Buffer (black)	790 µL	Room temperature
BS Reaction Buffer (black)	160 µL	Room temperature
BS Binding Buffer	4.8 mL	Room temperature
BS Wash Buffer	1 mL	Room temperature
BS Desulphonation Buffer	1600 µL	Room temperature
BS Elution Buffer (white)	176 µL	Room temperature
BS Spin Columns	8 columns	Room temperature
BS Collection Tubes	8 tubes	Room temperature
WGBS Primer Mix (green)	16 µL	-20°C
MethylTaq Plus 2X Master Mix (green)	200 µL	-20°C

Required Materials Not Provided

- Gloves to wear at all steps
- Ethanol 100% (room temperature)
- Nuclease-free water
- 8-tube strips or 0.2 mL PCR tubes
- Agencourt AMPure XP 5 mL (Beckman Coulter Genomics, Cat # A63880))
- DiaMag02 Magnetic Rack (Diagenode, Cat #B04000001)
- Heat block
- Thermocycler
- 2, 10, 20, 200 and 1000 µL pipettes
- Nuclease-free barrier pipette tips
- Microcentrifuge
- 1.5 mL nuclease-free microcentrifuge tubes
- Low melt agarose such as Low Gelling Temperature Agarose with a melt point of 65ºC (Boston Bioproducts, Cat # P-730)
- 1X TAE buffer
- Clean razor or scalpel
- SYBR Gold (Invitrogen, Cat # S11494)
- UV transilluminator or gel documentation instrument
- Gel electrophoresis apparatus

Equipments required for quality control:

- Fluorescence-based assay for DNA concentration measurement, e.g. the Qubit High Sensitivity assay (Life Technologies Cat #Q32851)
- Library analysis assay such as Agilent High Sensitivity DNA Kit for BioAnalyzer (Agilent, Cat #5067-4626)

Remarks before starting

Warnings and precautions

- DTT in buffers may precipitate after freezing. If precipitate is seen, vortex buffer for 1-2 minutes or until the precipitate is in solution. The performance of the buffer is not affected once precipitate is in solution.
- Do not heat the WGBS Adapters above room temperature.
- If starting with a DNA input amount greater than or less than 1 µg, adjust the WGBS Adapter volume to preserve the insert to adapter ratio.

Starting Material

The Premium WGBS Kit has been optimized using fragmented DNA.

Genomic DNA must be randomly sheared by sonication to generate fragments around 200 bp (see example below). All the DNA shearing protocols using Diagenode's sonication devices are available on our website: **www.diagenode.com**.

Example of shearing using the Bioruptor® Pico for WGBS:

The genomic DNA was diluted in TE buffer to reach a concentration of 100 ng/µL and 100 µL were sheared in a 0.65 mL Bioruptor® Microtube (Cat. No. C30010011).

The following program was used:

- Cycles: [30 seconds "ON" & 30 seconds "OFF"]
- 13 cycles

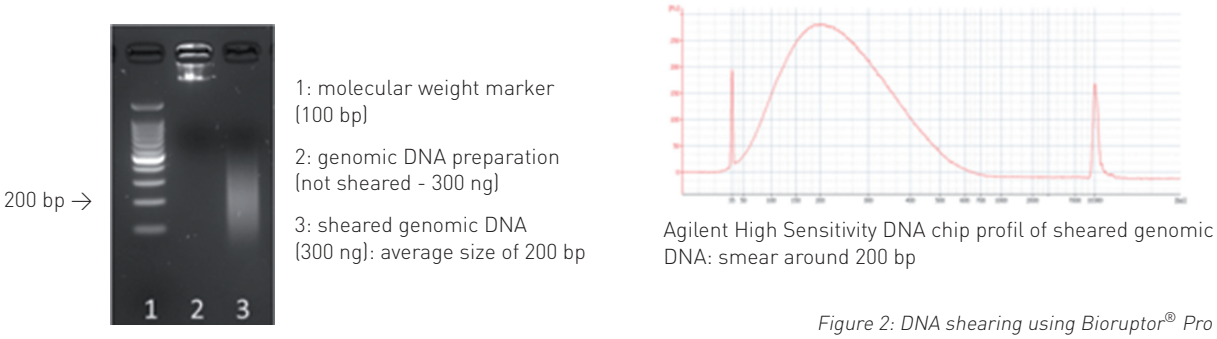

Figure 2: DNA shearing using Bioruptor® Pro

The Diagenode's Premium WGBS Kit has also been validated for ChIP-Bis-sequencing experiments. Thus the starting material can be some immunoprecipitated DNA after ChIP experiments against histone marks. For ChIP experiments we recommand the iDeal ChIP-seq kit for Histones (Cat. No. C01010051).

Starting inputs from 5 ng to 1 µg of fragmented genomic DNA or ChIP'd DNA are compatible with this kit.

Reagent Preparation

- Allow Agencourt AMPure XP Beads to come to room temperature and vortex the beads until liquid appears homogenous before every use.

Protocol

STEP 1. End Repair

1. For each sample, combine the following reagents on ice in a 8-tube strip or in a 0.2 mL PCR tube:

_ µL	Nuclease-free Water (white cap)
_ µL	Fragmented or ChIP'd genomic DNA (5 ng - 1 µg)
7 µL	WGBS End Repair Buffer Mix (clear cap)
3 µL	WGBS End Repair Enzyme Mix (clear cap)
50 µL	TOTAL

2. Mix thoroughly by pipetting.

3. Incubate on a thermocycler for 30 minutes at 22°C.

STEP 2. Clean-Up

1. Add 90 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.

2. Incubate at room temperature for 5 minutes.

3. Place the 8-tube strip on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.

4. Remove and discard clear supernatant. Take care not to disturb beads. Some liquid may remain in the tube.

5. Add 200 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.

6. Repeat step 5, for a total of 2 ethanol washes. Ensure all ethanol has been removed.

7. Remove the tube from the DiaMag02 and let dry at room temperature for 3 minutes.

8. Resuspend dried beads with 17 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the tube.

9. Incubate resuspended beads at room temperature for 2 minutes.

10. Place tube on DiaMag02 at room temperature for 5 minutes or until the sample appears clear.

11. Transfer 16 µL of clear sample to new tube.

12. If you wish to pause your experiment, the procedure may be safely stopped at this step and samples stored at -20°C. To restart, thaw frozen samples on ice before proceeding

STEP 3. Adenylation

1. Combine the following reagents in a 0.2 mL tube or 8-tube strip:

16 µL	End-Repaired DNA (from Step 2)
4.5 µL	WGBS Adenylation Mix (red cap)
20.5 µL	TOTAL

2. Mix thoroughly by pipetting.
3. Incubate on a thermocycler for 30 minutes at 37°C.

STEP 4: Adapter Ligation

If starting with less than 1 µg of input DNA, follow the 'Table 2 below to dilute the methylated adapters. This is important for retaining the optimal input:adapter ratio.

Table 2. Adapter Dilution

Input DNA	10 ng	100 ng	250 ng	500 ng
WGBS Adapter	1 µL	1 µL	1 µL	2 µL
WGBS Adapter Dilution Buffer	49 µL	9 µL	3 µL	2 µL
Amount per reaction	2.5 µL	2.5 µL	2.5 µL	2.5 µL

1. For each sample, combine the following reagents (in this order) in the PCR tube or 8-tube strip:

20.5 µL 3'	Adenylated DNA (from Step 3)
31.5 µL	WGBS Ligation Mix (purple cap)
2.5 µL	WGBS Adapter (purple cap)
54.5 µL	TOTAL

2. Mix thoroughly by pipetting.
3. Incubate on a thermocycler for 15 minutes at 22°C.

STEP 5. Clean-Up and Size Selection

There are two size selection protocol options to choose from:

- **Option 1** is a completely gel-free protocol that utilizes a magnetic bead based cleanup to size select DNA insert fragments between 100 – 400 bp. If you would like to avoid gel steps and are interested in an insert size between 100 – 400 bp, follow option 1.
- **Option 2** utilizes a magnetic bead based clean-up followed by agarose gel size selection and is designed for users interested in DNA insert fragments →130 bp. The user can size select a specific range of DNA fragments post-ligation. To follow option 2 go directly to page 13.

Option 1: Gel-free Size Selection Clean-Up

1. Add 44 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.

2. Incubate at room temperature for 5 minutes.

3. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.

4. Remove and discard clear supernatant. Take care not to disturb beads.

5. Add 200 µL of freshly prepared 80% ethanol to each sample and incubate plate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.

6. Repeat step 5, for a total of 2 ethanol washes and ensure all ethanol has been removed.

7. Remove the tube from the DiaMag02 and let dry at room temperature for 3 minutes.

NOTE:
Do not over dry the beads.

8. Resuspend dried beads with 52 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the well.

9. Incubate resuspended beads at room temperature for 2 minutes.

10. Place tube on DiaMag02 at room temperature for 5 minutes or until the sample appears clear.

11. Transfer 50 µL of clear sample to new tube.

12. Add 25 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.

13. Incubate at room temperature for 5 minutes.

14. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.

15. Do not discard clear sample in this step. Transfer 73 µL of clear sample to a new tube.

16. Add 14 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting

17. Incubate at room temperature for 5 minutes.

18. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.

19. Remove and discard clear supernatant. Take care not to disturb beads.

20. Add 200 µL of freshly prepared 80% ethanol to each sample and incubate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.

21. Repeat step 20, for a total of 2 ethanol washes and ensure all ethanol has been removed.

22. Remove the tube from the DiaMag02 and let dry at room temperature for 3 minutes

NOTE:
Do not over dry the beads.

23. Resuspend dried beads with 22 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the tube.

24. Incubate resuspended beads at room temperature for 2 minutes.

25. Place plate on DiaMag02 at room temperature for 5 minutes until the sample appears clear.

26. Transfer 20 µL of clear sample to new tube.

27. If you wish to pause your experiment, the procedure may be safely stopped at this step and samples stored at -20°C. To restart, always thaw your frozen samples on ice before proceeding.

Option 2: Clean-Up and Agarose Gel Size Selection

1. Add 55 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.
2. Incubate at room temperature for 5 minutes.
3. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.
4. Remove and discard clear supernatant. Take care not to disturb beads. Some liquid may remain in tubes.
5. Add 200 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate plate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.
6. Repeat step 5, for a total of 2 ethanol washes and ensure all ethanol has been removed.
7. Remove the plate from the DiaMag02 and let dry at room temperature for 3 minutes.
8. Resuspend dried beads with 52 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the well.
9. Incubate resuspended beads at room temperature for 2 minutes.
10. Place plate on DiaMag02 for 5 minutes or until the sample appears clear.
11. Transfer 50 µL of clear sample to new tube.
12. Add 55 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.
13. Incubate at room temperature for 5 minutes.
14. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.
15. Remove and discard clear supernatant. Take care not to disturb beads. Some liquid may remain in tubes.
16. Add 200 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate plate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.
17. Repeat step 16, for a total of 2 ethanol washes and ensure all ethanol has been removed.
18. Remove the tube from the DiaMag02 and let dry at room temperature for 3 minutes.
19. Resuspend dried beads with 22 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the tube.

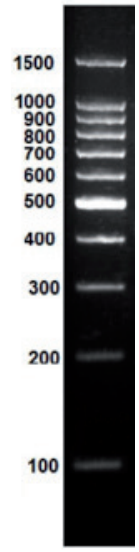

Figure 3. MW Ladder Ready-to-Load 100 bp

20. Incubate resuspended beads at room temperature for 2 minutes.

21. Place tube on DiaMag02 for 5 minutes or until the sample appears clear.

22. Transfer 20 µL of clear sample to new tube.

NOTE:
If you wish to pause your experiment, the procedure may be safely stopped at this step and samples stored at -20°C. To restart, thaw frozen samples on ice before proceeding.

23. Add 6 µL of 6X Gel Loading Dye (orange cap) to each sample.

24. Prepare pre-stained SYBR Gold 2% low melt agarose gel by adding 15 µL of SYBR Gold to every 150 mL of cooled 1X TAE and agarose gel buffer. Mix and then pour into gel tray. Load the entire sample into one lane of the gel. If processing more than one sample, it is recommended to run separate gels or leave several empty wells between samples to avoid cross contamination.

25. Load 4 µL of MW Ladder Ready-to-Load 100 bp (orange cap) into one lane, skipping at least two lanes between it and your sample.

26. Run the gel with 1X TAE buffer at 100-120V for 60 -120 minutes.

27. Visualize the gel on a UV transilluminator or gel documentation instrument (see figure 2).

28. Use a clean razor or scalpel to cut out a slice of gel from each sample lane corresponding to the 250-300 bp marker or other reference control. This results in an insert size of130-180 bp (WGBS DNA Adapters add ~120 bp to each fragment). The user may choose other insert sizes when appropriate. Keep in mind that sequence reads that overlap into the adapter will result in reads that do not map to the reference sequence.

29. Add 400 µL of DNA Binding Buffer to each gel slice containing sample and mix well.
Incubate your sample at room temperature and vortex the sample occasionally until the agarose is completely melted. For larger gel slices, cut smaller pieces and use 3-5 mL of DNA Binding Buffer.

30. For every 400 µL of binding buffer, add 20 µL of 100% ethanol to each sample and mix well.

31. Transfer the sample to a Clean-Up Spin Column.

32. Centrifuge the Clean-Up Spin Column in a microcentrifuge at 14,000 rpm for 1 minute.

33. Decant the flow through and return the Clean-Up Spin Column into the same collection tube.

34. Repeat steps 31-33 until all of the sample has been processed.

35. Add 700 µL of DNA Wash Buffer to each column.

36. Centrifuge the Clean-Up Spin column in a microcentrifuge at 14,000 rpm for 1 minute.

37. Decant the flow through and return the Clean Up Spin Column into the same collection tube

38. Repeat steps 35 -37 one time.

39. Centrifuge the Clean-Up Spin column in a microcentrifuge at 14,000 rpm for 1 minute to remove any residual ethanol.

40. Place the Clean-Up Spin Column into a clean 1.5 mL nuclease-free microcentrifuge tube. Add 22 µL of Column Elution Buffer (yellow cap) to the center of the column. Incubate the column at room temperature for 1 minute.

41. Centrifuge the Clean-Up Spin Column in a microcentrifuge at 14,000 rpm for 1 minute to elute the clean DNA. If you wish to pause your experiment, the procedure may be safely stopped at this step and samples stored at -20°C. To restart, thaw frozen samples on ice before proceeding.

STEP 6. Bisulfite conversion

1. Before the first use add 4 mL of 100 % ethanol to 1 mL BS Wash Buffer concentrate. The BS Conversion Reagent supplied within this kit is a solid mixture and must be prepared prior to first use.

2. Add 790 µL of BS Solubilization Buffer (black cap) and 300 µL of BS Dilution Buffer (black cap) to the tube of BS Conversion Reagent.

3. Mix at room temperature with frequent vortexing or shaking for 10 minutes.

4. Add 160 µL of BS Reaction Buffer (black cap) and mix an additional 1 minute.

NOTE:
It is normal to see trace amounts of undissolved reagent in the BS Conversion Reagent. Each tube of BS Conversion Reagent is designed for 8 separate DNA treatments including an excess.

STORAGE: The BS Conversion Reagent is light sensitive, so minimize its exposure to light. For best results, the BS Conversion Reagent should be used immediately following preparation. If not used immediately, the BS Conversion Reagent solution can be stored overnight at room temperature, one week at 4°C, or up to one month at -20°C. Stored BS Conversion Reagent solution must be warmed to 37°C, then vortexed prior to use.

5. Add 117 µL of BS Conversion reagent to 20 µL of sample from step 5, mix, centrifuge briefly and incubate in a thermocycler as indicated in Table 3.

Table 3

Cycle Step	Temperature	Time	Cycles
Denaturation	95°C	1 minute	20
Conversion	60°C	10 minutes	
Hold	4°C	∞	

6. Place BS Spin columns into the provided BS Collection Tubes.

7. Add 600 µL of BS Binding Buffer into a BS Spin Column.

8. Load bisulfite-converted libraries into the BS Spin Columns containing the BS Binding Buffer. Close the cap and mix by inverting the column several times.

9. Centrifuge at full speed (>10,000 x g) for 30 seconds. Discard the flow-through.

10. Add 100 µL of BS Wash Buffer to each column. Centrifuge at full speed for 30 seconds.

11. Add 200 µL of BS Desulphonation Buffer to each column and let stand at room temperature (20-30°C) for exactly 30 minutes. After the incubation, centrifuge at full speed for 30 seconds.

12. Add 200 µL of BS Wash Buffer to each column. Centrifuge at full speed for 30 seconds. Add another 200 µL of BS Wash Buffer and centrifuge for an additional 30 seconds.

13. Place each column into a 1.5 mL tube. Add 17 µL of BS Elution Buffer (white cap) directly to the center of the column matrix and wait for 2 minutes. Centrifuge for 30 seconds at full speed to elute the DNA.

14. Elute one more time by transferring the eluate to the column's membrane. Wait 2 minutes, then centrifuge for 30 seconds at full speed.

NOTE:
Proceed with the PCR amplification reaction quickly.

STEP 7: PCR Amplification

The table 4 can serve as a guideline in deciding the number of PCR cycles.

Table 4

Input DNA	5-10 ng	100 ng	250 ng	500 ng	1 µg
Fragmented DNA	18 cycles	14-17 cycles	14-17 cycles	11-14 cycles	9-11 cycles

1. For each sample, combine the following reagents (in this order) in the PCR tube or 8-tube strip:

_ µL	Bisulfite Converted Product (from Step 6)
_ µL	Nuclease free Water (white cap)
25 µL	MethylTaq Plus 2X Master Mix (green cap)
2 µL	WGBS Primer Mix (green cap)
50 µL	TOTAL

2. Mix thoroughly by pipetting.

3. PCR Cycles

10 min	95°C	
30 sec	98°C	Repeat 9 - 18 cycles**
30 sec	65°C	
45 sec	72°C	
7 min	72°C	

**PCR cycles will vary depending on the amount of starting material and quality of your sample. Further optimization may be necessary. Always use the least number of cycles possible.

4. Add 55 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.

5. Incubate at room temperature for 5 minutes.

6. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.

7. Remove and discard clear supernatant. Take care not to disturb beads. Some liquid may remain in tubes.

8. Add 200 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate plate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.

9. Repeat step 8, for a total of 2 ethanol washes and ensure all ethanol has been removed.

10. Remove the tube from the DiaMag02 and let dry at room temperature for 3 minutes.

11. Resuspend dried beads with 52 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the tube.

12. Incubate resuspended beads at room temperature for 2 minutes.

13. Place tube on DiaMag02 for 5 minutes or until the sample appears clear.

14. Transfer 50 µL of clear sample to new tube.

15. Add 55 µL of AMPure XP Beads to each sample and mix thoroughly by pipetting.
16. Incubate at room temperature for 5 minutes.
17. Place the tube on the DiaMag02 at room temperature for 5 minutes or until the supernatant appears clear.
18. Remove and discard clear supernatant. Take care not to disturb beads. Some liquid may remain in tubes.
19. Add 200 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.
20. Repeat step 19, for a total of 2 ethanol washes and ensure all ethanol has been removed.
21. Remove the tube from the DiaMag02 and let dry at room temperature for 3 minutes.
22. Resuspend dried beads with 11.5 µL of Resuspension Buffer. Mix thoroughly by pipetting. Ensure beads are no longer attached to the side of the well.
23. Incubate resuspended beads at room temperature for 2 minutes.
24. Place tube on DiaMag02 for 5 minutes or until the sample appears clear.
25. Transfer 10 µL of clear sample to a new tube.
26. Examine your library with a fluorometer and check the size using an Agilent Bioanalyzer or 2% agarose gel (See Examples in Figure 4).

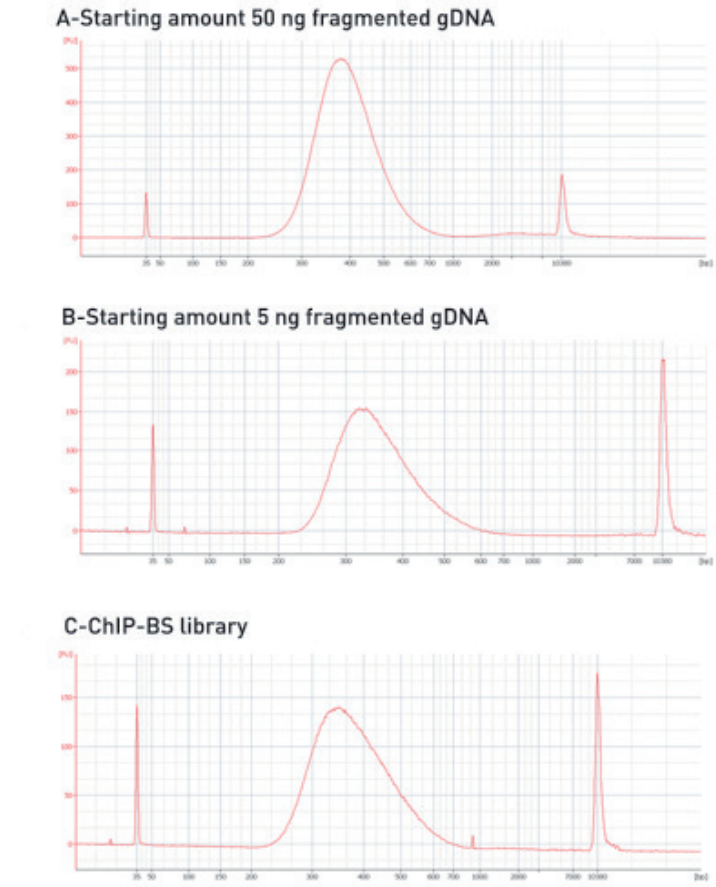

Figure 4. Bioanalyzer profile of different libraries obtained with the Premium WGBS Kit.

A. **50 ng** of fragmented genomic DNA was used. A total of 18 cycles of PCR were performed after bisulfite conversion. 1 µL of the resulting library was diluted 5-fold and run on an Agilent High Sensitivity DNA chip.

B. **5 ng** of fragmented genomic DNA was used. A total of 18 cycles of PCR were performed after bisulfite conversion. 1 µL of the resulting library was run on an Agilent High Sensitivity DNA chip.

C. **50 ng** of ChIP'ed DNA was used. The Chromatin Immunoprecipitation was performed against the H3K27me3 histone mark, using the Diagenode's iDeal ChIP-seq Kit for Histones (Cat #C01010050 and C01010051). A total of 18 cycles of PCR were performed after bisulfite conversion. 1 µL of the resulting library was diluted 2.5-fold and run on an Agilent High Sensitivity DNA chip.

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