

H2A.Zac antibody

Cat. No. C15410173

Lot:	A1775P
Size:	10 µg / 50 µg
Type:	Polyclonal, ChIP-grade, ChIP-seq grade, CUT&Tag-grade
Source:	Rabbit
Concentration:	1.4 µg/µl

Specificity:	Human: positive Other species: not tested
Purity:	Affinity purified polyclonal antibody
Storage buffer:	PBS containing 0.05% azide and 0.05% ProClin 300

Storage: Store at -20°C; for long storage, store at -80°C. Avoid multiple freeze-thaw cycles.

Precautions: This product is for research use only. Not for use in diagnostic or therapeutic procedures.

Description: Polyclonal antibody raised in rabbit against histone H2A.Z acetylated at lysines 4, 7 and 11, using a KLH-conjugated synthetic peptide.

Applications

Applications	Suggested dilution	References
ChIP/ChIP-seq*	0.5 µg per ChIP	Fig 1, 2
CUT&Tag	1 µg	Fig 3
ELISA	1:5,000	Fig 4
Dot blotting	1:20,000	Fig 5
Western blotting	1:1,000	Fig 6
Immunofluorescence	1:500	Fig 7

*Please note that the optimal antibody amount per IP should be determined by the end-user. We recommend testing 0.5-5 µg per IP.

Target description

Histones are the main constituents of the protein part of chromosomes of eukaryotic cells. They are rich in the amino acids arginine and lysine and have been greatly conserved during evolution. Histones pack the DNA into tight masses of chromatin. Two core histones of each class H2A, H2B, H3 and H4 assemble and are wrapped by 146 base pairs of DNA to form one octameric nucleosome. Histone tails undergo numerous post-translational modifications, which either directly or indirectly alter chromatin structure to facilitate transcriptional activation or repression or other nuclear processes. In addition to the genetic code, combinations of the different histone modifications reveal the so-called "histone code". Histone methylation and demethylation is dynamically regulated by respectively histone methyl transferases and histone demethylases. Acetylation of the histone H2A variant H2A.Z is associated with the promoters of active genes.

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Results

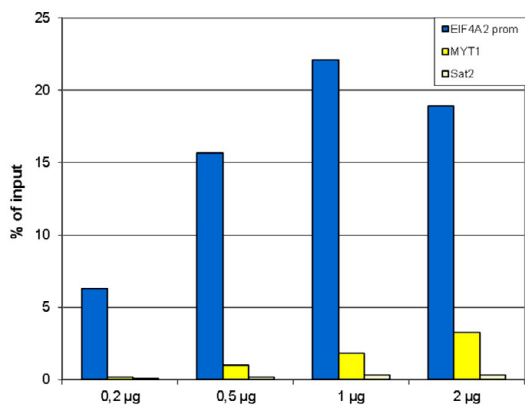


Figure 1: ChIP results obtained with the antibody directed against H2A.Zac

ChIP assays were performed using HeLa cells, the antibody against H2A.Zac (cat. No. C15410173) and optimized primer pairs for qPCR. ChIP was performed on sheared chromatin from 100,000 K562 cells using the iDeal ChIP-seq kit. A titration of the antibody consisting of 0.2, 0.5, 1 and 2 μ g per ChIP experiment was analysed. IgG (1 μ g/IP) was used as negative IP control. QPCR was performed using primers specific for the promoter of the EIF4A2 gene, used as positive control target and for the coding region of the MYT1 gene, and the Sat2 satellite repeat, used as negative control targets. Figure 1 shows the recovery (the relative amount of immunoprecipitated DNA compared to input DNA).

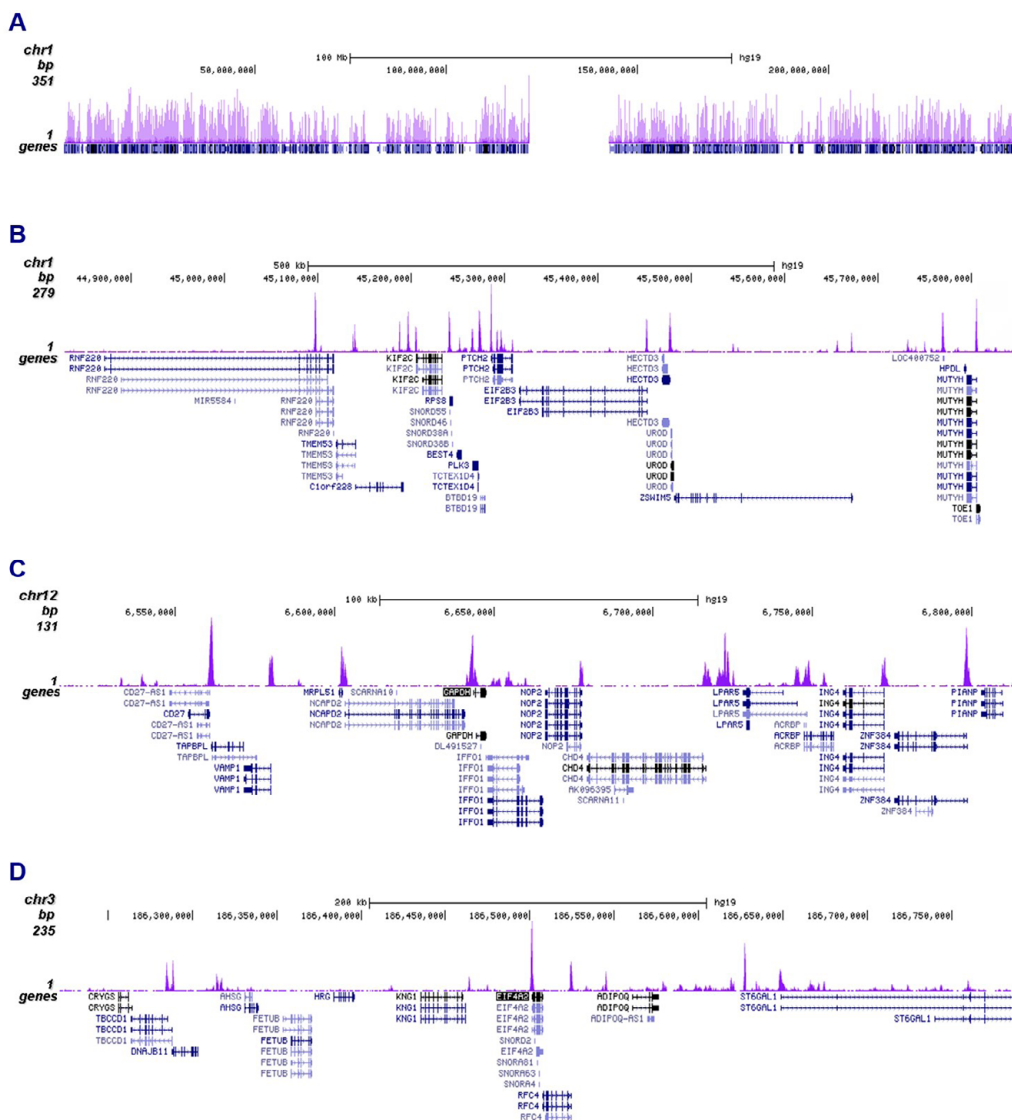


Figure 2: ChIP-seq results obtained with the antibody directed against H2A.Zac

ChIP was performed with 0.5 µg of the antibody against H2A.Zac (cat. No. C15410173) as described above. The IP'd DNA was subsequently analysed with an Illumina Genome Analyzer. Library preparation, cluster generation and sequencing were performed according to the manufacturer's instructions. The 36 bp tags were aligned to the human genome using the ELAND algorithm. Figure 2 shows the peak distribution along the complete sequence and a 1 Mb region of human chromosome 1 (figure 2A and B) and in two regions surrounding the GAPDH and the EIF4A2 positive control gene (figure 2C and D, respectively).

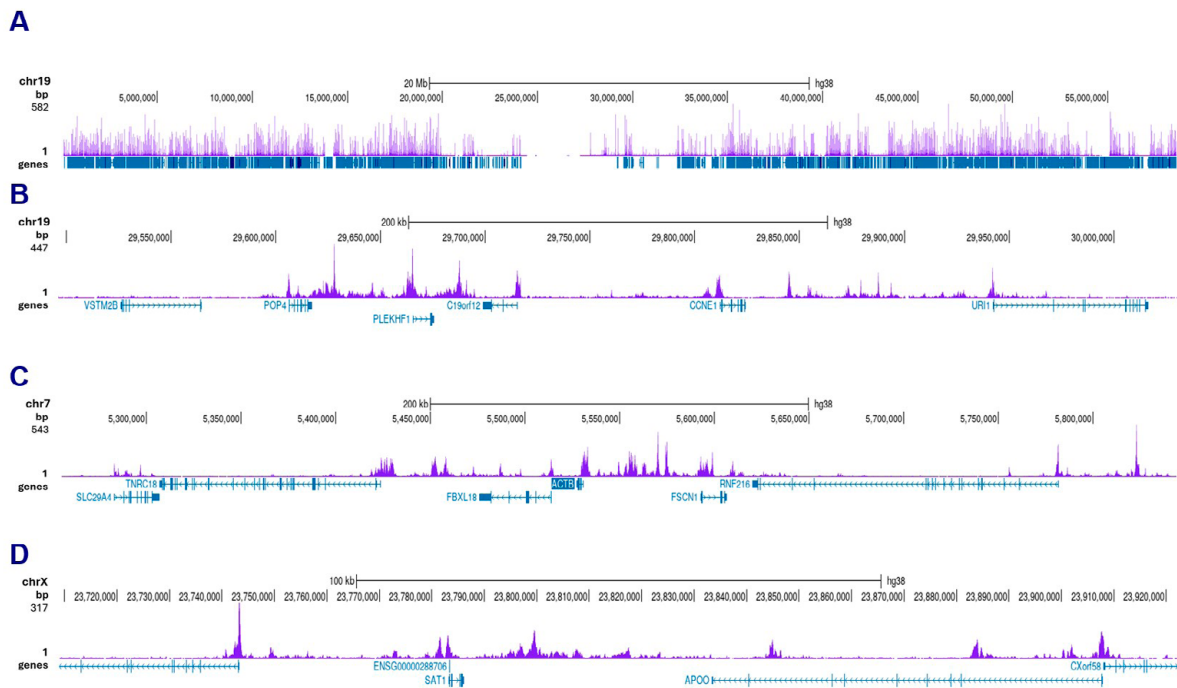


Figure 3: CUT&Tag results obtained with the antibody directed against H2A.Zac

CUT&Tag was performed on 50,000 K562 cells using 1 µg of the antibody against H2A.Zac (cat. No. C15410173) and the Universal CUT&Tag kit (cat. No. C01070024). The libraries were subsequently analysed on an Illumina NovaSeqX sequencer (2x150 bp paired-end reads) according to the manufacturer's instructions. The tags were aligned to the human genome (hg38) using the BWA algorithm. Figure 3 shows the peak distribution along the complete sequence and a 600 kb region of chromosome 19 (figure 3A and B) and in 2 genomic regions on chromosome 7 and X (figure 3C and D, respectively).

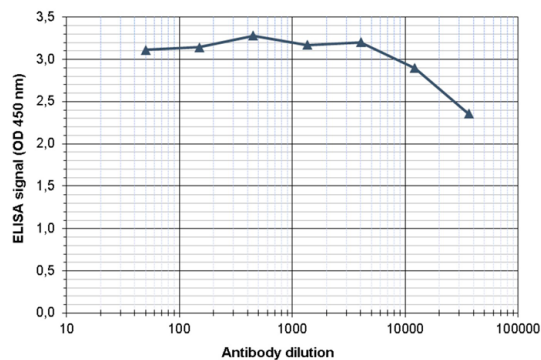


Figure 4: Determination of the antibody titer

To determine the titer of the antibody, an ELISA was performed using a serial dilution of the antibody directed against H2A.Zac (cat. No. C15410173). The antigen used was a peptide containing the histone modifications of interest. By plotting the absorbance against the antibody dilution (Figure 4), the titer of the purified antibody was estimated to be 1:265,000.

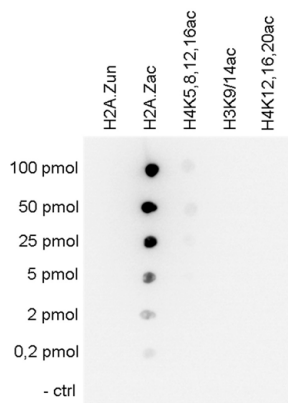


Figure 5: Cross reactivity test using the antibody directed against H2A.Zac

A Dot Blot analysis was performed to test the cross reactivity of the antibody against H2A.Zac (cat. No. C15410173) with peptides containing other histone acetylations and the unmodified H2A.Z sequence. One hundred to 0.2 pmol of the respective peptides were spotted on a membrane. The antibody was used at a dilution of 1:20,000. Figure 5 shows a high specificity of the antibody for the modification of interest.

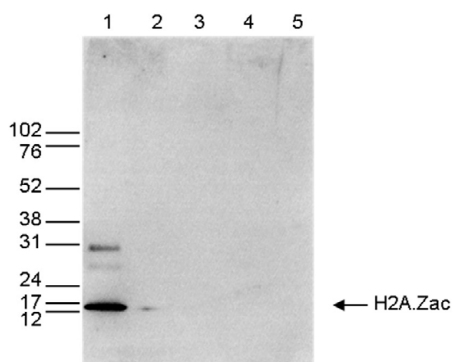


Figure 6: Western blot analysis using the antibody directed against H2A.Zac

Western blot was performed on whole cell extracts (25 µg, lane 1) from HeLa cells, and on 1 µg of recombinant histone H2A, H2B, H3 and H4 (lane 2, 3, 4 and 5, respectively) using the antibody against H2A.Zac (cat. No. C15410173). The antibody was diluted 1:1,000 in TBS-Tween containing 5% skimmed milk. The position of the protein of interest is indicated on the right, the marker (in kDa) is shown on the left.

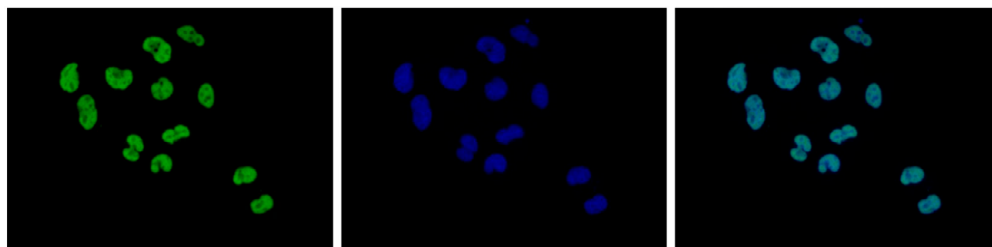


Figure 7: Immunofluorescence using the antibody directed against H2A.Zac

HeLa cells were stained with the antibody against H2A.Zac (cat. No. C15410173) and with DAPI. Cells were fixed with 4% formaldehyde for 10' and blocked with PBS/TX-100 containing 5% normal goat serum and 1% BSA. The cells were immunofluorescently labelled with the H2A.Zac antibody (left) diluted 1:500 in blocking solution followed by an anti-rabbit antibody conjugated to Alexa488. The middle panel shows staining of the nuclei with DAPI. A merge of the two stains is shown on the right.