

H3K4me1 antibody

Cat. No. C15200150

Lot:	001-12
Size:	50 µg
Type:	Monoclonal, ChIP-grade
Isotype:	NA
Source:	Mouse
Concentration:	1 µg/µl

Specificity:	Human: positive.
	Other species: not tested
Purity:	Protein A purified monoclonal antibody

Storage buffer: PBS containing 0.05% azide.

Storage: Store at -20°C; for long storage, store at -80°C. Avoid multiple freeze-thaw cycles.

Precautions: This product is for research use only. Not for use in diagnostic or therapeutic procedures.

Description: Monoclonal antibody raised in mouse against histone H3, monomethylated at lysine 4 (H3K4me1), using a KLH-conjugated synthetic peptide.

Applications

Applications	Suggested dilution	References
ChIP*	1 µg per ChIP	Fig 1
ELISA	1:3,000	Fig 2
Western blotting	1:1,000	Fig 3
Immunofluorescence	1:500	Fig 4

*Please note that the optimal antibody amount per IP should be determined by the end-user. We recommend testing 1-5 µg per IP.

Target description

Histones are the main constituents of the protein part of chromosomes of eukaryotic cells. They are rich in the amino acids arginine and lysine and have been greatly conserved during evolution. Histones pack the DNA into tight masses of chromatin. Two core histones of each class H2A, H2B, H3 and H4 assemble and are wrapped by 146 base pairs of DNA to form one octameric nucleosome. Histone tails undergo numerous post-translational modifications, which either directly or indirectly alter chromatin structure to facilitate transcriptional activation or repression or other nuclear processes. In addition to the genetic code, combinations of the different histone modifications reveal the so-called "histone code". Histone methylation and demethylation is dynamically regulated by respectively histone methyl transferases and histone demethylases.

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Results

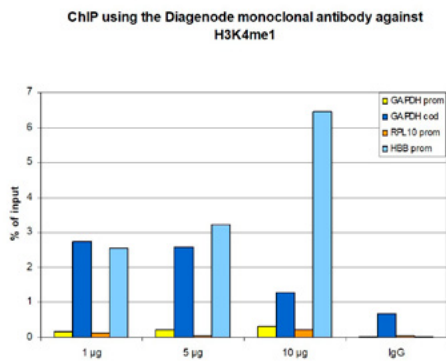


Figure 1: ChIP results obtained with the Hologic Diagenode monoclonal antibody directed against H3K4me1

ChIP assays were performed using HeLa cells, the monoclonal antibody against H3K4me1 (cat. No. C15200150) and optimized PCR primer sets for qPCR. ChIP was performed with the “iDeal ChIP-seq kit for Histones (cat. No. C01010051), using sheared chromatin from 1 million cells. A titration of the antibody consisting of 1, 5 and 10 µg per ChIP experiment was analysed. IgG (5 µg/IP) was used as negative IP control. QPCR was performed with primers for the promoter and the coding region of the GAPDH gene, and for the RPL10 and HBB promoters. Figure 1 shows the recovery, expressed as a % of input (the relative amount of immunoprecipitated DNA compared to input DNA after qPCR analysis).

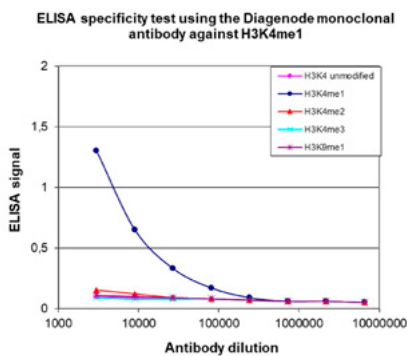


Figure 2: Cross reactivity of the Hologic Diagenode monoclonal antibody directed against H3K4me1

To test the specificity an ELISA was performed using a serial dilution of the Hologic Diagenode monoclonal antibody against H3K4me1 (cat. No. C15200150). The wells were coated with peptides containing the unmodified H3K4 as well as the mono-, di- and trimethylated H3K4 and the monomethylated H3K9. Figure 2 shows a high specificity of the antibody for the modification of interest.

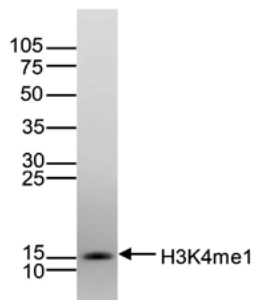


Figure 3: Western blot analysis using the Hologic Diagenode monoclonal directed antibody against H3K4me1

Histone extracts (15 µg) from HeLa cells were analysed by Western blot using the Hologic Diagenode monoclonal antibody against H3K4me1 (cat. No. C15200150) diluted 1:1,000 in TBS-Tween containing 5% skimmed milk. The position of the protein of interest is indicated on the right; the marker (in kDa) is shown on the left.

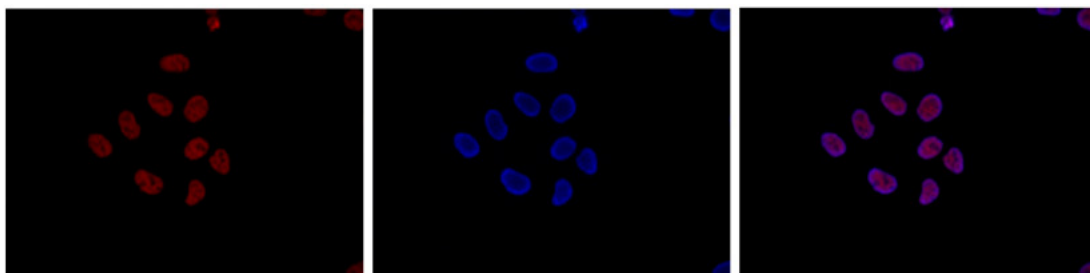


Figure 4: Immunofluorescence using the Hologic Diagenode monoclonal antibody directed against H3K4me1

HeLa cells were stained with the Diagenode antibody against H3K4me1 (cat. No. C15200150) and with DAPI. Cells were fixed with 4% formaldehyde for 10' and blocked with PBS/TX-100 containing 5% normal goat serum and 1% BSA. The cells were immunofluorescently labelled with the H3K4me1 antibody (left) diluted 1:500 in blocking solution followed by an anti-mouse antibody conjugated to Alexa594. The middle panel shows staining of the nuclei with DAPI. A merge of the two stainings is shown on the right.